

MICROBIOLOGY DEPARTMENT

STANDARD OPERATING PROCEDURE		
Department: Microbiology	SOP No.:	
Title: Microbiological Assay for Antibiotic & Vitamin B ₁₂	Effective Date:	
Supersedes: Nil	Review Date:	
Issue Date:	Page No.:	

1.0 **OBJECTIVE**

To lay down a procedure for operation for Microbial assay.

RESPONSIBILITY 2.0

Microbiologist/QC Executive

3.0 **ACCOUNTABILITY**

Quality Control Manager

4.0 **PROCEDURE**

4.1 INTRODUCTION

4.1.1 The inhibition of microbial growth under standardized condition may be utilized for demonstrating the therapeutic efficacy of antibiotics. Any such changes in the antibiotics molecules which may not be defected by chemicals methods will be revealed by a change in the antimicrobial activity and microbiological assays are very useful for resolving doubts regarding possible change in potency of antibiotics and their preparation. The microbial assay is bases upon a comparison of the inhibition of growth of micro-organisms by known concentration of a standard preparation of antibiotics having known activity.

4.2 **METHOD: CUP PLATE METHOD**

4.2.1 The Cup-Plate methods depends upon diffusion of antibiotics from a vertical cylinder through a solidified agar layer in a petri-dish or plate to an extent such that growth of the added microorganisms is prevented entirely in a zone around the cylinder containing solution of antibiotics.

4.3 REQUIRMENTS

4.3.1 Given Antibiotics to be assayed, working standard, dehydrated media, Buffer solution, Initial solvents, Test Organism, Spectrophotometer, sterile petri-dish, sterile volumetric flask& cylinder (25 ml).

PREPARATION OF STANDARD SOLUTION 4.4



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4.4.1 To prepare a stock solution, dissolve a quantity of the standard preparation of a given antibiotics, accurately weight; indicated in the table–3 (Appendix 9.1/Indian Pharmacopoeia, Volume –2) in the solvent specified in the table, and then dilute to the required concentration as indicated.

4.5 PREPARATION OF THE SAMPLE SOLUTION

4.5.1 From the information available for the substances being examined at (the unknown) assign to it and an assumed potency per unit weight or volume, and on this assumption prepare on the day of the assay a stock solution and test dilution as specified for each antibiotics in table 3 (Appendix 9.1/Indian Pharmacopoeia Volume –2). But with the name final diluent as used for the standard preparation.

4.6 TEST ORGANISMS.

4.6.1 The test organism for each antibiotics is listed in Table 4 (Appendix 9.1/Indian Pharmacopoeia, Volume –2) together with it identification number (ATCC Number). Maintain a culture on slant of medium and under the incubation condition specified in Table 5 (Appendix 9.1/Indian Pharmacopoeia, Volume –2) and transfer weekly to fresh slants.

4.7 PREPARATION OF INNOCULUM

4.7.1 Take the test organism maintained on the slant, and scrap its growth with Nichrome wire loop with 25 ml sterile 0.9% Saline. Incubate for 30 minutes and determine the dilution factor which will give 25% light transmission at about 530 nm. Determine the amount of suspension to be added to each 100 ml agar.

4.8 PREPARATION OF PLATE

- 4.8.1 Inoculate the previously liquidified medium appropriate to the assay with require quantity of suspension of micro-organism, add the suspension to the medium at a temperature between 40 to 50°C and immediately pour the inoculated medium into petri-dishes to give a depth of 3-4 mm. Ensure that the layers of medium are uniform in thickness, by placing the petri-dishes on a level surface.
- 4.8.2 The prepared dishes must be stored in a manner so as to ensure that no significant growth or death of the organism occurs before the petri-dishes are used for Assay.
- 4.8.3 Using the appropriate buffer solution indicated in table 2 & 3 (Appendix 9.1/Indian Pharmacopoeia, Volume –2). Prepare solutions



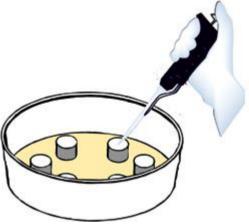
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of known concentration of the standard preparation and solutions of the corresponding assumed concentration of the antibiotic to be examined . Apply the solutions to the surface of the solid medium in well (Well in the agar surface should be made by punching into the agar surface with the help of SS316L borer). The volume of the solution added to each well must be uniform and sufficient almost fill the wells (i.e.100 $\mu l)$.







Pipetting dilution

- 4.8.4 When the petri-dishes are used, arrange the solutions of the standard preparation and the antibiotic to be examined on each dish so that they alternate around the dish and so that the highest concentration of the standard and test preparation are not adjacent.
- 4.8.5 Leave the petri-dishes are used, arrange the solutions of the standard preparation and the antibiotics to be examined on each dish so that they alternate around the dish and so that the highest concentration of the standard and test preparation are not adjacent. Leave the petri-dishes standing for about 1 hrs. for pre-incubation diffusion. Incubate the petri-dishes at 37°C for 18-24 hours.

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After the incubation period, accurately measure the diameters of zones 4.8.6 of inhibition and calculate the results.







Antibiotic Zone of Inhibition

Measuring Antibiotic Zone Vitamin B₁₂ Zone of Exhibition

4.9 ESTIMATION OF POTENCY

(TH + TL) - (SH + SL)a =(TH + TL) - (SH + SL)

 $M = a \times I$

Potency of Sample =potency ratio x dilution factor x units per mg of standard

SH Standard high Concentration SL Standard Low Concentration TH Test high Concentration **Test Low Concentration** TLM $log Potency ratio = a \times I$ log Ratio of dosage Potency Ratio -Antilog M

5.0 REASON FOR REVISION

New SOP.

7.0 **DISTRIBUTION:**

Certified Copy No. 1 : Head of Department – Quality Control

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8.0 ANNEXURES:

Annexure-I : Format for Microbial Assay for Vancomycin Hydrochloride

Annexure-II : Format for Microbial Assay for Vitamin B₁₂

9.0 **REFERENCES**:

Pharmaceutical Microbiology Manual", FDA, 2014

Indian Pharmacopoeia



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	MICROBIOLOGIC	CAL ASSAY FOR V	NEXURE-I ANCOMYCIN HYDRO PPLATE METHOD)	OCHLORIDE STERILE USP	
Product/Item	Name:		_		
Batch No.:				tibiotic Medium No. 8 M 041	
Batch Size:				sms: Bacillus subtilis (ATCC 66	533)
Mfg Date:			Sampled On		
Exp Date: Date of testing	α•		Sampled By Released On		
Potency of the Standard Weig	Distilled water Final of Std. =	mcg/mg $mg/100 ml \rightarrow 10 ml/s$	2 ml/25 ml 2 ml/25 ml 2 ml/10 2 ml/25 0 ml 2 ml/10	00 ml. 5 ml	
Plate No.	TH	TL	SH	SL	
01 02					
03					
04 Avg.(mm)					
CALCULATION a = (TH+T) (TH-T) I = Log of dose M= Log Poten	ON OF POTENCY: (L) - (SH+SL) (L) + (SH-SL) es: 0.6021 cy ratio = a x 0.6021 =	===		e, SL: Std. low dose	
Potency ratio =	= Antilog M =				
Potency of san	nple = Potency ration	x dilution factor x Un	its/ ml. of std.		
	=	mcg/mg. or	n as is basis.		

ANALYSED BY/DATE CHECKED BY/DATE



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ANNEXURE-II	
ANNEAURE-II	

MICROBIOLOGICAL ASSAY FOR VITAMIN B₁₂ (BY CUP PLATE METHOD)

Batch No. :	Medium B ₁₂ Culture agar (E. coli Maintenance Medium)
	(E. coli Mutant Culture Agar)
Batch Size :	Test Organisms: Escherichia coli, (ATCC 11105)
Mfg Date :	Sampled on :
Exp Date :	Sampled By :
Date of testing :	Released on :
Testing Details:	

Potency of the Std. = _____mcg/mg

Std Wt.: _____ mg/50 ml \rightarrow 5 ml/100 ml. \rightarrow 10/100 ml \rightarrow 10/100 ml \rightarrow 5 ml/25 ml. (SH)(0.25conc.) (SL) 0.05 conc.

Test wt. taken: $\underline{\qquad}$ mcg/50 ml ---- 5 ml/25 ml

(TH) 0.25conc. (TL) 0.5conc

Plate	TH	TL	SH	SL
01				
02				
03				
04				
Avg.(mm)				

Key: TH: Test high dose, TL: Test low dose SH: Std. High dose, SL: Std. low dose

CALCULATION OF POTENCY:

(TH+TL) - (SH+SL)
a = = =
(TH-TL) + (SH-SL) I = Log of doses: 0.699 (fixed value)
M= Log Potency ratio = a x I ==
Potency ratio = Antilog M =
Potency of sample = Potency ration x dilution factor x Units/ ml. of std.

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