

MICROBIOLOGY DEPARTMENT

### **EFFICACY OF FUMIGATION**

1.	Unit Operation: To ensure that the fumigation is effective in reducing the bioload of the working environment.					
2.	Objective: To determine that the fumigation procedure effectively reduces the bio-load of					
3.	Site of Study: Microbiology laboratory					
4.	Validation Team: Representatives from	Quality Control  Quality Assurance	:			
5.	Area to be validated:					
6.	SOP for the study:  i. SCA, SCDA plates are prepared as per  ii. Plate exposure done as per  iii. Fumigation and defumigation done as per					
<b>7.</b> a) b)	Control: Pre-incubated Soyabean Casein Digest agar plates and Sabouraud's chloramphenicol agar plates are used. Unexposed spore strip of <i>Bacillus subtilis</i> var niger ATCC 9372.					
8.	Materials and Equipments Used: Prepare the media from dehydrated media using purified water. Test the media for pH, Growth promotion ability and pre-incubation.					
9.	Experimental Results: Carry out the experiment as per validation protocol.					



MICROBIOLOGY DEPARTMENT

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10. OBSERVATIONS: I)		
Date of fumigation:	Date of De-fumigation:	
Residual formaldehyde content:		
A) Time of plate exposure before fum	Date/sign:	
Incubation started at:hrs.		Incubator No.
Location	Bacterial cfu/plate before fumigation Load no.	Fungal count before fumigation Load no.
Date /sign		
Incubation completed at: hrs.		
B) Time of plate exposure after defum	Date/sign:	
Incubation started at: hrs		Incubator No.
Location	Bacterial cfu/plate after fumigation Load no.	Fungal cfu/plate after fumigation Load no.
Date /sign Incubation completed at: hrs.		



MICROBIOLOGY DEPARTMENT

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II) Viable count of biological Indicators:	
Viable count of biological Indicators (unexposed strips)	
Viable count of biological Indicators after fumigation (exposed strips)	
11. Conclusion:	
12. Recommendation:	
13. Team approval:	
Quality Control	Quality Assurance
Date:	
14. Review (inclusive of follow up action, if any):	
15. Approved by:	
QUALITY ASSURANCE Date:	UNIT HEAD



MICROBIOLOGY DEPARTMENT

#### **EFFICACY OF FUMIGATION**

#### 1. Unit Operation:

To ensure that the fumigation is effective in reducing the bioload of the working environment

#### 2. Objective:

To determine that the fumigation procedure effectively reduces the bio-load of Microbiology laboratory.

#### 3. Site of Study:

Microbiology Laboratory

#### 4. Validation Team:

Representatives from: Quality Control

Quality Assurance

(Individuals to be named in validation report.)

#### 5. SOP for the study:

- i. SCA, SCDA plates are prepared as per SOP.
  - ii Environment count by settle plate technique as per SOP.
  - iii Fumigation and de-fumigation of the area as per SOP.

#### 6. Control:

- c) Pre-incubated Soyabean Casein Digest agar plates and Sabouraud's chloramphenicol agar plates are used.
- d) Unexposed spore strip of Bacillus subtilis var niger ATCC 9372.

#### 7. Materials and Equipments Used:

Prepare the media from dehydrated media using purified water. Test the media for pH, Growth promotion ability and pre-incubation.

#### 8. Experimental details:

- a) Prepare the Soyabean casein digest agar and Sabouraud chloramphenicol agar and pour plates as given. Pre-incubate the plates for 48 hours to check for contamination.
- b) Carry out the environmental control by settle plate technique in the micro testing area before fumigating the areas as per the SOP.
- c) Expose the at least three spore strips of Bacillus subtilis var niger ATCC 9372 in sterile petri plate before starting the fumigation activity in the testing area.
- d) Carry out fumigation and defumigation of the area as per the procedure given.
- e) After defunigation check for residual formaldehyde content and then carryout the environmental control by settle plate technique in micro testing area as per SOP.
- f) Incubate the Soyabean casein digest agar plates at 30 to 35°C for 72 hours and Sabouraud's chloramphenicol agar at 20 to 25°C for 5 days in the inverted position.
- g) Count the number of colonies observed and record it in the test data sheet.



MICROBIOLOGY DEPARTMENT

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h) Carry out the viable count of exposed and unexposed biological indicator strips as per SOP.

#### 9. Acceptance Criteria

- a) The environment count in the area should show reduction in bacterial and fungal count after fumigation
- b) Formaldehyde content in the area after defumigation for I hour should not be more than 1 ppm
- c) There should be reduction in the viable count of exposed spore strips as compared to the unexposed strips.

COMPILED BY	APPROVED BY	AUTHORISED BY	
Unit - Quality Assurance	Corporate - Quality Assurance	Unit head	
Date:	Date :	Date:	