

PHARMA DEVILS MICROBIOLOGY DEPARTMENT

PROTOCOL FOR EVALUATION OF EFFECTIVENESS OF DISINFECTANT & SANITATION SOLUTION (BY MOPPING METHOD)

Protocol for Evaluation of Effectiveness of Disinfectant & Sanitation Solution (By Mopping method)



MICROBIOLOGY DEPARTMENT

PROTOCOL FOR EVALUATION OF EFFECTIVENESS OF DISINFECTANT & SANITATION SOLUTION (BY MOPPING METHOD)

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1.0 PROTOCOL APPROVAL:

Signing of this Approval page of Validation Protocol indicates agreement with the Validation

approach described in this document will be prepared and approved.

If any modification in the validation approach becomes necessary, an addendum shall be Prepared.

	NAME	SIGNATURE/DATE
Prepared By: Officer, Quality Control		
Reviewed By: Executive, Quality Control		
Reviewed By: Head, Quality Control		
Reviewed By: Head, Regulatory Affairs		
Approved By: Head, Factory		
Approved By: Head, Quality		



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- 2.0 **PURPOSE:** The purpose of this document to provides methodology for evaluation of effectiveness of sanitation solutions against the microorganisms by mopping method.
- 3.0 **SCOPE:** This protocol is applicable to all sanitation solutions used for sanitation of production area and QC microbiology lab.

4.0 **PROCEDURE:**

Activity	Sign
For Bacteria Streak a loopful of microorganisms on to the surface of Soyabean Casein Digest Agar Media, Lot No Incubation: Incubate at 30-35°C for 24 to72 hrs. Incubator NoDate/Time In Date/Time Out	
Observe for growth.	
For Fungi Streak a loopful of microorganisms on to the surface of Sabouraud Dextrose agar Media, Lot No Incubation: Incubate at 20-25°C for 72 to120 hrs. Incubator NoDate/Time In Date/Time Out Observe for growth.	
Harvest the growth in test tube containing of 9 ml of sterile Normal saline for Bacteria and sterile Normal saline for yeast and fungi.	
Vortex the test tubes for about 30 seconds to homogenate the contents.	
Make serial dilution up to 10 ⁻⁸ using sterile Normal saline for Bacteria and sterile Normal saline for yeast and fungi.Vortex to homogenate the contents for about 30 seconds while preparing each dilution.	



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Record the details	in the below	table.						
Name of Organism	Date of Serial Dilution /Incubation	Dilution	Volume Tested	Count /I Plate 1	Plate Plate 2	Avg.	Final Inoculum Population	Observed By (Sign / Date)
Bacillus subtilis								
Escherichia coli								
Staphylococcus aureus								
Candida albicans								
Aspergillus niger								
Environmental isolate								
duplicate.								
duplicate. Add about 15 to 20 of melted media ju medium for bacter with the agar and a) ml of previ st prior to ac ia/ Sabourau	ously melt ldition in t id Dextros	ed and co o plates) se agar N	ooled to and reco Aedium	at not m ord the te for yeas	ore that emperation that the second s	n 45°C (mea ture. Soyabe fungi, mix th	le petriplates in asure the temperatur can casein digest aga ne culture suspension organism and cultur
duplicate. Add about 15 to 20 of melted media ju medium for bacter with the agar and a dilution.) ml of previ st prior to ac ia/ Sabourau llow it to so	ously melt ldition in t ıd Dextros lidify at ro	eed and co o plates) se agar M om tempo	ooled to and reco Medium erature.Io	at not m ord the te for yeas dentify y	ore that emperant t and f with nat	an 45°C (mea ture. Soyabe fungi, mix th ame of micro	asure the temperature ean casein digest age ne culture suspension
duplicate. Add about 15 to 20 of melted media ju medium for bacter with the agar and a dilution. Repeat above step) ml of previ st prior to ac ia/ Sabourau llow it to so for remainin	ously melt ldition in t ıd Dextros lidify at ro ıg three dil	eed and co o plates) se agar M om tempo ution i.e.	ooled to and reco Medium erature.Io	at not m ord the te for yeas dentify y	ore that emperant t and f with nat	an 45°C (mea ture. Soyabe fungi, mix th ame of micro	asure the temperature ean casein digest age ne culture suspension
of melted media ju medium for bacter) ml of previ st prior to ac ia/ Sabourau llow it to so for remainin edure for rer	ously melt ldition in t id Dextros lidify at ro g three dil naining or	eed and co o plates) se agar M om tempo ution i.e. ganisms.	ooled to and reco Aedium erature.Io 10 ^{-6,} 10	at not m ord the te for yeas dentify y	ore that emperant t and f with nat	an 45°C (mea ture. Soyabe fungi, mix th ame of micro	asure the temperature ean casein digest age ne culture suspension
duplicate. Add about 15 to 20 of melted media ju medium for bacter with the agar and a dilution. Repeat above step Repeat above proc Immediately store) ml of previ st prior to ac ia/ Sabourau llow it to so for remainin edure for rer the culture s	ously melt ldition in t id Dextros lidify at ro ig three dil naining or uspension	eed and co o plates) se agar M om tempo ution i.e. ganisms. s in refrig	ooled to and reco Aedium erature.Io 10 ^{-6,} 10	at not m ord the te for yeas dentify y	ore that emperant t and f with nat	an 45°C (mea ture. Soyabe fungi, mix th ame of micro	asure the temperature ean casein digest age ne culture suspension
duplicate. Add about 15 to 20 of melted media ju medium for bacter with the agar and a dilution. Repeat above step Repeat above proc) ml of previ st prior to ac ia/ Sabourau llow it to so for remainin edure for rer the culture s	ously melt ldition in t id Dextros lidify at ro ig three dil naining or uspensions n inverted	eed and co o plates) se agar M om tempo ution i.e. ganisms. s in refrig	ooled to and reco Aedium erature.Io 10 ^{-6,} 10	at not m ord the te for yeas dentify y	ore that emperate t and t with nation 0 ⁻⁸ as v	an 45°C (mea ture. Soyabe fungi, mix th ame of micro	asure the temperature ean casein digest age ne culture suspension



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Observe the plates and record number of cfu/plate.

Preparation of sanitation solution :

Prepare in use concentration of sanitation solution using purified water (PW) and mixwell. Aseptically filter the sanitation solution using a 0.45 micron membrane filter under a laminar air flow (LAF) and collect the filtered sanitation solution in a sterile container.

Main procedure :

Select the surface (approx 5x5 or 10x10Cms) and make grid for the sampling in such way to prevent the repetative position and sampling.sanitized surface with approved sanitation solution

. Take one RODAC and label it as blank and incubate as below table.

Media Lot No. ____

_____ Observation : ______ cfu/plate.

Add 1 ml of culture suspension having the concentration of selective dilution .spread the culture suspension on surface and allow it to air dry.

Take five samples by RODAC plates from the five different positions from surface (initial) to check the actual bioburden, label accordingly and incubate the plates. Media lot No. _____

Take sterile sponge sheet or any other, deep it into the filtered sanitation solution and mope the surface and note down time of mopping started ______.

At different time intervals of contact period, take 5 samples by RODAC plate from the different surfaces and note down the time of sampling, label accordingly and incubate.

For Bacteria	For Fungi
Incubate at 30-35°C for 24 to 72 hrs	Incubate at 20-25°C for 72 to 120 hrs
Incubator No.:	Incubator No.:
Date/Time in :	Date/Time in :
Date/Time out:	Date/Time out:



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Observe for CFU and record in the table given. Name of Disinfectant: **Date of Test: Concentration: Date of Observation:** Name of Initial Dilution Population Observed count after contact time Log reduction observed organism Popula Selected Taken for (X-Y) **(Y)** Test CFU tion 0 Min 5 Min 10 Min 15 Min 0 Min 5 Min 10 Min 15 Min Observed /ml (X) CFU by (Sign /Date) /ml **Bacillus** subtilis Escheric hia coli Staphylo coccus aureus Candida albicans Aspergill us niger Environ mental isolate **ACCEPTANCE CRITERIA:** There should be at least three log reduction of initial count achieved within 15 minutes of contact time.

CONCLUSION :

The disinfectant.....is qualified as per above specification.

Done by: Sign/Date: Checked By: Sign/Date:



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5.0 **REVISION HISTORY:**

S.No.	Revision No.	Reason for Revision	Effective date
1	00	First-edition – Hence Not applicable	