



STANDARD OPERATING PROCEDURE

Department: Microbiology

SOP No.:

Title: Bacterial Endotoxin Test (LAL)

Effective Date:

Supersedes: Nil

Review Date:

Issue Date:

Page No.:

1.0 OBJECTIVE:

To lay down a procedure for Bacterial Endotoxin Test.

2.0 SCOPE:

This SOP is applicable for Bacterial Endotoxin Test in Microbiology Section of Quality Control Area.

3.0 RESPONSIBILITY:

Officer/Executive – Microbiologist

4.0 ACCOUNTABILITY:

Head – QC

5.0 PROCEDURE:

5.1 The Bacterial Endotoxin Test (BET) is a test to detect or quantify endotoxins from gram –negative bacteria using ameobocyte lysate from the horseshoe crab (*Limulus polyphemus* or *Tachypleus tridentatus*). There are three techniques for this test: The Gel clot technique, which is based on gel formation, The turbidimetric technique, based on the development of turbidity after cleavage of an endogenous substrate, and the chromogenic technique, based on the development of color after cleavage of a synthetic peptide-chromogen complex.

5.2 Receive the LAL Reagent through supplier and make an entry of receipt of reagent in Titled “**Lysate Receipt and Consumption Record**” in Respective **Annexure-III**,

5.3 MATERIAL AND INSTRUMENTS:

5.3.1 Limulus Amebocyte Lysate Reagent.

5.3.2 Control Standard Endotoxin (CSE).

5.3.3 LAL Reagent Water.

5.3.4 Depyrogenated Dilution Tubes.

5.3.5 Depyrogenated LAL Assay Tube (10 x 75 mm).

5.3.6 Calibrated Micropipette, Pyrogen free micro tips (20 – 200 µl).



STANDARD OPERATING PROCEDURE

Department: Microbiology

SOP No.:

Title: Bacterial Endotoxin Test (LAL)

Effective Date:

Supersedes: Nil

Review Date:

Issue Date:

Page No.:

5.3.7 Calibrated Micropipette, Pyrogen free micro tips (100 – 1000 µl).

5.3.8 Vortex Mixer.

5.3.9 Heating Block.

5.3.10 Depyrogenated Test Tube (18 x 120 mm).

5.4 Switch “ON” the Heating Block and set the Temperature at 37° C ± 1°C.

5.5 PREPARATION OF CONTROL STANDARD ENDOTOXIN (CSE):

5.5.1 Receive the Control Standard Endotoxin (CSE) through supplier and make an entry of receipt in Titled “**Control Standard Endotoxin (CSE) Receipt and Consumption Record**” in Respective **Annexure–VII**, and Reconstitute CSE in LRW as per Manufacturer’s Instruction.

5.5.2 Use Reconstituted CSE as per Manufacturer’s Instruction after Reconstitutions; store Reconstituted CSE at 2°C – 8°C Temperature or as per Manufacturer’s Instructions.

5.5.3 Prepare further dilutions as given below (in case of 100 EU/vial).

S. No.	Endotoxin	LRW	Endotoxin Concentration (EU/ml)
1.	100 EU/vial	5 ml	20 EU/ml
2.	0.1 ml of 20EU/ml	0.9 ml	2 EU/ml
3.	0.5 ml of 2EU/ml	0.5 ml	1 EU/ml (8λ)
4.	0.5 ml of 1EU/ml	0.5 ml	0.5 EU/ml (4λ)
5.	0.5 ml of 0.5EU/ml	0.5 ml	0.25 EU/ml (2λ)
6.	0.5 ml of 0.25EU/ml	0.5 ml	0.125 EU/ml (λ)
7.	0.5 ml of 0.125EU/ml	0.5 ml	0.06EU/ml (λ/2)
8.	0.5 ml of 0.06EU/ml	0.5 ml	0.03 EU/ml (λ/4)
9.	0.5 ml of 0.03 EU/ml	0.5 ml	0.015 EU/ ml (λ/8)
10.	0.5 ml of 0.015 EU/ml	0.5 ml	0.007 EU/ ml (λ/16)

5.5.4 The supplied COA is specific for the lysate lot and CSE lot. So use the same lot for testing.

5.6 RECONSTITUTION OF LYSATE:



STANDARD OPERATING PROCEDURE

Department: Microbiology	SOP No.:
Title: Bacterial Endotoxin Test (LAL)	Effective Date:
Supersedes: Nil	Review Date:
Issue Date:	Page No.:

- 5.6.1** Now reconstitute the lysate by opening the aluminum seal. Collect lysate powder into the bottom of the vial by tapping on a hard surface and then open the cap slowly.
- 5.6.2** As per manufacturer's instruction, add LRW to lysate vial by avoiding direct contact with fingers and close the cap immediately. Do not vortex lysate.
- 5.6.3** Reconstituted lysate shall be stored at 2°C – 8°C in refrigerator and to be used within 24 hrs. of reconstitution.

5.7 CONFIRMATION OF THE LABELED LYSATE SENSITIVITY:

- 5.7.1** Confirmation of the lysate sensitivity must be carried out when a new batch of lysate is use or when there is any change in the experimental conditions which may affect the outcome of the test or any alteration in test result. Confirm in four replicates the labeled sensitivity λ , expressed in EU/ml or IU/ml, of the lysate solution prior to use in the test.
- 5.7.2** Take 20 Depyrogenated Assay Tubes and label the tubes by numbering and arrange quadruplicate in stand and proceed the test as per mentioned below:

Tubes	CSE Dilution Used	LRW	Lysate in μ l	No. of Replicates
2 λ	100 μ l of 2 λ	–	100 μ l	4
λ	100 μ l of λ	–	100 μ l	4
$\lambda/2$	100 μ l of $\lambda/2$	–	100 μ l	4
$\lambda/4$	100 μ l of $\lambda/4$	–	100 μ l	4
Negative water control (NWC)	–	100 μ l	100 μ l	4

- 5.7.3** Pipette 100 μ l diluted CSE i.e. to 2 λ , λ , $\lambda/2$ and $\lambda/4$ separately into depyrogenated assay tubes. For NC use 100 μ l of LRW.
- 5.7.4** Add 100 μ l of reconstituted lysate into each tube.
- 5.7.5** Incubate the tubes in heating block at 37°C \pm 1°C for 60 \pm 2 minutes, avoiding vibration.
- 5.7.6** After incubation, take each tube and invert through approximately 180° in one smooth motion. If a firm gel has formed that remains in place upon inversion, record the result as positive. A result is negative if an intact gel is not formed.



STANDARD OPERATING PROCEDURE

Department: Microbiology	SOP No.:
Title: Bacterial Endotoxin Test (LAL)	Effective Date:
Supersedes: Nil	Review Date:
Issue Date:	Page No.:

5.7.7 The test is not valid unless the lowest concentration of the standard solutions shows a negative result in all replicate tests.

5.7.8 The endpoint is the last positive result in the series of decreasing concentrations of endotoxin. Calculate the mean value of the logarithms of the end-point concentrations and then the antilogarithm of the mean value using the following expression.

$$\text{Geometric mean end - point concentration} = \text{antilog} \frac{\sum e}{f}$$

$\sum e$ = sum of the log end-point concentrations of
the dilution series used,
 f = number of replicates.

5.7.9 The geometric mean end-point concentration is the measured sensitivity of the lysate solution (EU/ml or IU/ml). The Lysate sensitivity should not less than $\lambda/2$ and not more than 2λ .

5.7.10 Record the Consumption Details of Lysate in Titled “Lysate Receipt And Consumption Record” in Respective **Annexure-III**.

5.7.11 Record the details of Lysate Sensitivity in, Titled “Lysate Sensitivity Record” in Respective **Annexure- IV**.

5.8 CALCULATION FOR DETERMINING THE ENDOTOXIN LIMIT

5.8.1 The Endotoxin limit for active substances administered parenterally, Define on the Basis of Dose, is equal to

$$\text{Endotoxin Limit} = K/M$$

K = Threshold pyrogenic dose of Endotoxin per kilogram of body mass in a single hour period. 5 EU/kg body weight for parenteral drugs except those administered intrstheally. 0.2 EU/kg for intrathecal drugs.

M = Maximum recommended dose of product per Kilogram of body mass in a single hour period. (For this calculation, it is assumed that the average person weight 70 kg. if pediatric dose is higher, it shall be used in the calculation)

5.8.2 The Endotoxin limit of active substances administered parenterally is specified in Unit such as IU/ml, IU/mg, IU/unit of biological activity.



STANDARD OPERATING PROCEDURE

Department: Microbiology

SOP No.:

Title: Bacterial Endotoxin Test (LAL)

Effective Date:

Supersedes: Nil

Review Date:

Issue Date:

Page No.:

5.8.3 Record the Endotoxin Limit in Titled “**Determination of Endotoxin Limit**” in Respective **Annexure- VI**.

5.9 MAXIMUM VALID DILUTION (MVD):

5.9.1 The maximum valid dilution is the maximum allowable dilution of a sample at which the Endotoxin limit can be determined. Determine the MVD using the Following Formulae.

$$\text{MVD} = \frac{\text{Endotoxin Limit X Potency of Product}}{\lambda \text{ (Lysate Sensitivity)}}$$

5.10 TEST PROCEDURE:

5.10.1 Record the Sample Receiving details in Titled “**Sample Receiving and Analysis Record for Bacterial Endotoxin Test**” in Respective **Annexure-V**. Perform the test at MVD/2 or appropriate MVD.

5.10.2 Take 8 depyrogenated tubes and label the two tubes each as product, PPC, PC and NC. Arrange the tube in stand and label as per mentioned below:

Solution Description	LRW in μl	4λ (CSE) in μl	Product Dilution	Lysate in μl	No. of Replicates
Negative Product Control (NPC)	50	-	50	100	2
Positive Product Control (PPC)	-	50	50	100	2
Positive Water Control (PWC)	50	50	-	100	2
Negative Water Control (NWC)	100	-	-	100	2

5.10.3 Add 50 μl of LRW in product tube and PC, and 100 μl in NC. Immediately add 50 μl of sample, which is diluted at MVD or appropriate in sample tubes and PPC, and then add 50 μl of CSE that is diluted to 4 λ in a PPC and PC.

5.10.4 Finally add 100 μl of lysate in all tubes and incubate in heating block, where the temperature is maintained at 37 \pm 1 $^{\circ}\text{C}$ for 60 \pm 2 minutes



STANDARD OPERATING PROCEDURE

Department: Microbiology	SOP No.:
Title: Bacterial Endotoxin Test (LAL)	Effective Date:
Supersedes: Nil	Review Date:
Issue Date:	Page No.:

5.10.5 Record the result of WFI /Pure Steam in Titled “**Bacterial Endotoxin Test Report For WFI / Pure Steam**” in Respective **Annexure-I**,

5.10.6 Record the result of Product in Titled “**Bacterial Endotoxin Test Report**” in Respective **Annexure-II**.

5.10.7 BACTERIAL ENDOTOXIN TEST PROCEDURE FOR RUBBER BUNG

5.10.8 SAMPLE PREPARATION:-Prepare the sample solution by using LAL reagent water. If necessary, adjust the pH of the test solution so that the pH of the mixture of the lysate and the test solution falls within the pH range specified by the lysate reagent. This is usually applies to a product with a pH in the range of 6.0 to 8.0. The pH may be adjusted by the use of acid, base (prepared in LRW) or a suitable Buffer, as recommended by the lysate manufacturer. Buffer must be validated to be free of detectable Endotoxin and interfering factors

5.10.9 Take five rubber bungs in a Depyrogenated Test Tube (18X120 mm) / polybag (22X150 mm) add 4 ml LRW and vortex the tube for five to ten minutes.

5.10.10 After vortexing performed the test as per Point No 5.9.

5.10.11 BACTERIAL ENDOTOXIN TEST PROCEDURE FOR DEPYROGENATED VIALS

5.10.12 Take Ten Depyrogenated Vials and add appropriate volume of LRW as per size of vial after adding LRW vortex the vials for five to ten minutes.

5.11 After vortexing performed the test as per Point No 5.9.

5.12 PRECAUTIONS

5.13 All test tubes, which will be used for the test of BET must first be Depyrogenated.

5.14 Rehydrated CSE shall be stored as per vender COA instructions.

5.15 The supplied COA is specific for Lysate Lot & CSE Lot. So use the same lot for testing.

5.16 Do not vortex Lysate

5.17 Mcropipette should be calibrated.

5.18 RESULTS/ACCEPTANCE CRITERIA:



STANDARD OPERATING PROCEDURE

Department: Microbiology

SOP No.:

Title: Bacterial Endotoxin Test (LAL)

Effective Date:

Supersedes: Nil

Review Date:

Issue Date:

Page No.:

5.18.1 After incubation each tube is interpreted as either positive or negative, positive test indicates the formation of firm gel capable of maintaining its integrity when the test tube is inverted at 180°.

5.18.2 Negative test is characterized by the absence of gel or by the formation of a viscous mass, which does not hold when the tube is inverted at 1800.

5.18.3 The test is considered when both replicates PPC & PWC are positive and NWC is negative.

5.18.4 The preparation being examined complies with the test when a negative result is found for both replicates of sample.

5.18.5 When a positive result is found for both replicates of sample, it does not comply with the test.

5.18.6 Repeat the test if a positive result is found for one replicate of sample and a negative result is found in another replicate.

5.18.7 The preparation being examined complies with the test, if a negative result is found for both replicates sample in the repeat test.

5.18.8 When positive result observed on both the tubes of Test Preparation, Investigate the cause of its Failure by checking following parameters.

5.18.9 Check Product Dilution, CSE Dilution and Lysate Dilution and Storage Condition.

5.18.10 Check Sensitivity Record of Lysate Lot and matched CSE.

5.18.11 Check Heating Block Temperature and Calibration.

5.18.12 Check Micropipette Calibration.

5.18.13 Check pH of the solution.

6.0 REFERENCES:

USP-38 (<85> Bacterial Endotoxin Test)

7.0 ANNEXURES:



STANDARD OPERATING PROCEDURE

Department: Microbiology

SOP No.:

Title: Bacterial Endotoxin Test (LAL)

Effective Date:

Supersedes: Nil

Review Date:

Issue Date:

Page No.:

ANNEXURE No.	TITLE OF ANNEXURE	FORMAT No.
Annexure – I	Bacterial Endotoxin test report for wfi / pure steam	
Annexure - II	Bacterial Endotoxin test report	
Annexure – III	Lysate receipt and consumption record	
Annexure – IV	Lysate sensitivity record	
Annexure – V	Sample receiving and analysis record for bacterial Endotoxin test	
Annexure – VI	Determination of Endotoxin limit	
Annexure – VII	Control standard Endotoxin (CSE) receipt and consumption record	

ENCLOSURES: SOP Training Record

8.0 DISTRIBUTION:

- Controlled Copy No. 01 Head Corporate Quality Assurance
- Controlled Copy No. 02 Head Quality Control
- Master Copy Quality Assurance Department

9.0 ABBREVIATIONS:

SOP	Standard Operating Procedure
No.	Number
Ltd.	Limited
QA	Quality Assurance
QC	Quality Control
LRW	LAL Reagent Water
CSE	Control Standard Endotoxin
LAL	Limulus Amebocyte Lysate
CSE	Control Standard Endotoxin
µl	Micro Liter
ml	Mille Liter
NPC	Negative Product Control
PPC	Positive Product Control
NWC	Negative Water Control
PWC	Positive Water Control

10.0 REVISION HISTORY:



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STANDARD OPERATING PROCEDURE

Department: Microbiology

SOP No.:

Title: Bacterial Endotoxin Test (LAL)

Effective Date:

Supersedes: Nil

Review Date:

Issue Date:

Page No.:

CHANGE HISTORY LOG

Revision No.	Details of Changes	Reason for Change	Effective Date	Updated By
00	Nil	Nil		



PHARMA DEVILS

MICROBIOLOGY DEPARTMENT

STANDARD OPERATING PROCEDURE

Department: Microbiology	SOP No.:
Title: Bacterial Endotoxin Test (LAL)	Effective Date:
Supersedes: Nil	Review Date:
Issue Date:	Page No.:

Negative Water Control (NWC)	
Positive Water Control (PWC)	

+ve: Gel formation

-ve : No Gel formation

DILUTION TABLE:

Solution	Tube No.	Product Dilution	LRW	Control Standard Endotoxin	LAL Reagent	Total Volume
Negative Product Control (NPC)	01	50 μ L	50 μ L	–	100 μ L	200 μ L
	02	50 μ L	50 μ L	–	100 μ L	200 μ L
Negative Control (NC)	01	–	100 μ L	–	100 μ L	200 μ L
	02	–	100 μ L	–	100 μ L	200 μ L
Positive Control (PC)	01	–	50 μ L	50 μ L	100 μ L	200 μ L
	02	–	50 μ L	50 μ L	100 μ L	200 μ L
Positive Product Control (PPC)	01	50 μ L	–	50 μ L	100 μ L	200 μ L
	02	50 μ L	–	50 μ L	100 μ L	200 μ L

+ve: Gel formation

-ve : No Gel formation

Remark: The sample complies/ does not comply as per specification.

Microbiologist:

Date:

Checked By:

Date:



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MICROBIOLOGY DEPARTMENT

STANDARD OPERATING PROCEDURE

Department: Microbiology	SOP No.:
Title: Bacterial Endotoxin Test (LAL)	Effective Date:
Supersedes: Nil	Review Date:
Issue Date:	Page No.:

ANNEXURE-II BACTERIAL ENDOTOXIN TEST REPORT

Name of Product		A.R. No.	
Batch No.		Sample Quantity	
Mfg Date		Date of Receiving	
Exp. Date		Date of Analysis	
Shift		Date of Release	
Endotoxin Limit		Depyrogenation Cycle No.	
HAO ID No		Heating Block ID No	

REAGENT DETAILS

REAGENT DETAILS	LYSATE	CSE	LRW
Lot No.			
Sensitivity/Potency			
Date of Opening			
Use Before			
Expiry Date			
Manufacturer			

MVD =	MVD/ 2 Value =	
	MVD/4 Value =	

S. No.	Product Dilution	Product	LRW

S. No.	CSE Dilution	CSE Concentration	CSE Volume	LRW

Heating Block Temperature	37°C ± 1°C	Incubation Time	60 ± 2 Minutes
Incubation Started at		Incubation Completed at	

DILUTION TABLE:



PHARMA DEVILS

MICROBIOLOGY DEPARTMENT

STANDARD OPERATING PROCEDURE

Department: Microbiology

SOP No.:

Title: Bacterial Endotoxin Test (LAL)

Effective Date:

Supersedes: Nil

Review Date:

Issue Date:

Page No.:

Solution	Tube No.	Product Dilution (.....)	LRW	Control Standard Endotoxin (4 λ)	LAL Reagent	Total Volume	Observation
Negative Product Control (NPC)	01	50μL	50 μL	–	100μL	200μL	
	02	50μL	50 μL	–	100μL	200μL	
Negative Water Control (NWC)	01	–	100μL	–	100μL	200μL	
	02	–	100μL	–	100μL	200μL	
Positive Water Control (PWC)	01	–	50 μL	50 μL	100μL	200μL	
	02	–	50 μL	50 μL	100μL	200μL	
Positive Product Control (PPC)	01	50μL	–	50μL	100μL	200μL	
	02	50μL	–	50μL	100μL	200μL	

+ve: Gel formation

-ve: No Gel formation

Remark: The sample complies/ does not comply as per specification.

Microbiologist:

Checked By:

Date:

Date:



PHARMA DEVILS

MICROBIOLOGY DEPARTMENT

STANDARD OPERATING PROCEDURE

Department: Microbiology	SOP No.:
Title: Bacterial Endotoxin Test (LAL)	Effective Date:
Supersedes: Nil	Review Date:
Issue Date:	Page No.:

ANNEXURE-IV LYSATE SENSITIVITY RECORD

REAGENT DETAILS	LYSATE	CSE	LRW
Lot No.			
Sensitivity/Potency			
Date of Opening			
Use Before			
Expiry Date			
Manufacturer			

Instrument Details:

Micropipette ID No		Date of Calibration		Next Due Date	
Micropipette ID No		Date of Calibration		Next Due Date	
Heating Block ID No		Date of Calibration		Next Due Date	
HAO ID No		Date of Calibration		Next Due Date	

CSE Dilution:

S.No.	CSE Dilution	CSE Concentration	CSE Volume	LRW
Heating Block Temperature		37°C ± 1°C	Incubation Time	60 ± 2 Minutes
Incubation Started at			Incubation Completed at	

Observations:

Tube No.	Endotoxin Concentration						NWC	Test End Point EU/ml	Log of End Point	Geometric mean end point
	0.25	0.125	0.06	0.03	0.015	0.007				
1										
2										
3										
4										

+ ve = Gel Formation

- ve = No Gel Formation

Remarks: The lysate lot complies/does not comply for its sensitivity as mentioned.

Microbiologist:

Date:

Checked By:

Date:



PHARMA DEVILS

MICROBIOLOGY DEPARTMENT

STANDARD OPERATING PROCEDURE

Department: Microbiology

SOP No.:

Title: Bacterial Endotoxin Test (LAL)

Effective Date:

Supersedes: Nil

Review Date:

Issue Date:

Page No.:

ANNEXURE-VI DETERMINATION OF ENDOTOXIN LIMIT

Product Name		API	
Product Concentration		Test Concentration	
Rout of Administration	IM/IV/ Intrathecally	Product Safety Factor	
Threshold Pyrogenic Dose of Endotoxin		Maximum Recommended Dose	

Determination of Endotoxin Limit:

Determination of Test Concentration:

Determination of Product Safety Factor:

Final Endotoxin Limit:

Remarks: The Final Endotoxin Limit of _____ is _____ IU/EU/mg/ml.

Microbiologist:

Checked By:

Date:

Date:

