



PHARMA DEVILS

MICROBIOLOGY DEPARTMENT

STANDARD OPERATING PROCEDURE

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| Department: Microbiology | SOP No.: |
| Title: Bio-burden of Primary Packing Material | Effective Date: |
| Supersedes: Nil | Review Date: |
| Issue Date: | Page No.: |

1.0 PURPOSE:

1.1 To lay down a procedure for determining the bio burden (Total bacterial and fungal count) level of Primary packaging materials being used for packing of Tablets, Capsules, Liquids, Ointments and Creams.

2.0 SCOPE:

2.1 This procedure is applicable to Microbiology Laboratory.

3.0 RESPONSIBILITY:

3.1 Officer/Executive - Quality Control.

4.0 ACCOUNTABILITY:

4.1 Head –QC department.

5.0 PROCEDURE: (Frequency: First two consignments of each vendor per year)

5.1 Make entry of packing materials in microbial limit test log book as per SOP in Annexure-II. Carry out the test under LAF for all primary packing materials.

5.2 For Bottles/Caps (method used: membrane filtration method)

5.2.1 Sampling units: 10 bottles/caps

5.2.2 For bottles:

5.2.2.1 Pipette 10 ml of sterile 0.1% peptone water from the conical flask and transfer into one bottle and vortex for 05-10 seconds on a vortex mixer.

5.2.2.2 After vortexing transfer the 0.1% peptone water from each bottle into sterile filter assembly fixed on the manifold.

5.2.2.3 Pass 0.1% peptone water through 0.45 μ membrane filter by using vacuum pump.

5.2.2.4 Like wise repeat for the other four bottles as mentioned in 5.2.2.1 to 5.2.2.3.



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- 5.2.2.5 Discard the bottles after testing.
- 5.2.2.6 Carefully open the filter assembly. Ensure that the membrane filter is not sticking to the bottom of the filtration cup.
- 5.2.2.7 Aseptically remove the membrane filter from the assembly by using sterile forceps.
- 5.2.2.8 Place the membrane filter on to the surface of SCDA plate in such a way that the filtrate on the filter paper is facing upperside and the bottom is in direct contact with the media.
- 5.2.2.9 Incubate the above plate at 30°C -35°C for 3 days.
- 5.2.2.10 On completion of incubation period count the number of colonies observed on the membrane filter and record the total bacterial count as 'X' colony forming unit(cfu)/5 units in Annexure -I.
- 5.2.2.11 Repeat the above steps from 5.2.2.1 to 5.2.2.8 for fungal count by using Sabouraud dextrose agar (SDA) plate in place of SCDA and incubate the plate at 20°C -25°C for 5 days.
- 5.2.2.12 On completion of incubation period count the number of colonies observed on the membrane filter and record the total fungal count as 'Y' cfu/5 units in Annexure-I.
- 5.2.3 **For caps:**
- 5.2.3.1 Take two wide mouthed 250ml bottles containing 50 ml of sterile 0.1% peptone water. Place five No. of caps into each bottle.
- 5.2.3.2 Shake vigorously for one minute.
- 5.2.3.3 Open the mouth and pour the 0.1% peptone water into sterile filter assembly fixed on the manifold. Care shall be taken for the caps not to fall into the filtration cup.
- 5.2.3.4 Pass the solution through 0.45 μ membrane filter by using vacuum pump.
- 5.2.3.5 Discard the caps after testing.
- 5.2.3.6 Follow the steps from 5.2.2.6 to 5.2.2.12.
- 5.3 For Tubes Used For Ointments/Creams(method used: membrane filtration method)**
- 5.3.1 Sampling Units: 20 Tubes.



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- 5.3.2 Pipette 4 ml of sterile 0.1% peptone water from the conical flask and transfer into one tube from the open end, fold the open end and vortex for 05-10 seconds on a vortex mixer.
- 5.3.3 After vortexing transfer the 0.1% peptone water from the tube into sterile filter assembly fixed on the manifold. Pass 0.1% peptone water through 0.45 μ membrane filter by using vacuum pump.
- 5.3.4 Like wise repeat for the other tubes as mentioned in 5.3.1 and 5.3.2.
- 5.3.5 Discard the tubes after testing.
- 5.3.6 Carefully open the filter assembly. Ensure that the membrane filter is not sticking to the bottom of the filtration cup.
- 5.3.7 Aseptically remove the membrane filter from the assembly by using sterile forceps. Place the membrane filter on to the surface of SCDA plate in such a way that the filtrate on the filter paper is facing upperside and the bottom is in direct contact with the media.
- 5.3.8 Incubate the above plate at 30°C -35°C for 3 days.
- 5.3.9 On completion of incubation period count the number of colonies observed on the membrane filter and record the total bacterial count as 'X' colony forming unit(cfu)/5 units in Annexure -I.
- 5.3.10 Repeat the above steps from 5.3.2 to 5.3.7 for fungal count by using Sabouraud dextrose agar (SDA) plate in place of SCDA and incubate the plate at 20°C -25°C for 5 days.
- 5.3.11 On completion of incubation period count the number of colonies observed on the membrane filter and record the total fungal count as 'Y' cfu/5 units in Annexure-I.
- 5.4 For PVC Film / Aluminium Foil(method used: contact plate method)**
- 5.4.1 Carefully remove initial 2 to 3 meters of the foil/film from the roll and discard.
- 5.4.2 Open the lid of the Contact plate containing SCDA medium, bring the open portion of media in contact with the inner surface areas of the foil /film.
- 5.4.3 Gently press the plate with palm and hold, so that the media come in contact with the foil / film for 5-10 seconds.



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- 5.4.4 Slowly relax the palm and remove the plate from the contact. Take Care that the media should not stick to the foil / film.
- 5.4.5 Close the lid of the contact plate.
- 5.4.6 Cut off the tested portion of the foil / film and destroy.
- 5.4.7 Incubate the above plate at 30°C -35°C for 3 days.
- 5.4.8 On completion of incubation period count the number of colonies observed on SCDA plate and record the total bacterial count as `X' cfu/plate in Annexure-I.
- 5.4.9 Repeat the above steps from 5.4.1 to 5.4.6 for fungal count by using Sabouraud dextrose agar (SDA) plate in place of SCDA and incubate the plate at 20°C -25°C for 5 days.
- 5.4.10 On completion of incubation period count the number of colonies observed on the SDA plate and report the total fungal count as `Y' cfu/plate in Annexure-I.

Limits:

| Material | Total Bacterial Count | Total Fungal Count |
|------------------------------|-----------------------|-----------------------|
| Bottles | NMT 50 cfu / 5 units | NMT 10 cfu / 5 units |
| Caps | NMT 50 cfu / 5 units | NMT 5 cfu / 5 units |
| Tubes for ointments / creams | NMT 50 cfu / 10 units | NMT 10 cfu / 10 units |
| Aluminium foil / PVC film | NMT 20 cfu / plate | NMT 2 cfu / plate. |

ANNEXURE(S):

Annexure -I: Bio burden of primary packaging materials.

REFERENCES:

NIL.