

MICROBIOLOGY DEPARTMENT

STANDARD OPERATING PROCEDURE			
Department: Microbiology	SOP No.:		
Title: Environmental Monitoring of Microbiology Testing Area	Effective Date:		
Supersedes: Nil	Review Date:		
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1.0 OBJECTIVE:

To lay down a procedure for Environmental Monitoring of Microbiology testing area.

2.0 SCOPE:

This SOP is applicable for Environmental Monitoring in Microbiology testing area.

3.0 RESPONSIBILITY:

Microbiologist -Quality Control

Head-Quality Control

4.0 **PROCEDURE:**

4.1 Passive Air Sampling by Settle Plate Method:

- 4.1.1 Take 70% v/v IPA and lint free sterile mopping cloth.
- 4.1.2 Prepare Soybean casein digest agar medium as per SOP and mark the plates with Name of Medium, Lot No., Date of Preparation, Use before and Sign and date on bottom of plates and proceed for pre incubation not less than 48 hours at 20°–25°C.

4.2 Procedure for Plates Exposing:

- 4.2.1 Take required number of preincubated plates Keep the plates in petricanes, previously disinfect the petricanes with 70 % v/v IPA and keep in the dynamic pass box.
- 4.2.2 Follow the entry/exit procedure as per reference SOP for Microbiology testing area.
- 4.2.3 Write down the location number, location name, and initial time of exposure on SCDA plates and put the sterile SCDA plate on S. S. plate exposure stand of respective location.
- 4.2.4 Expose the petriplate at designated position by opening the lid of pre incubated SCDA plate.
- 4.2.5 Exposed the pre incubated SCDA plate for 04 hours + 15 minutes.
- 4.2.6 After completion of exposure period close the lid carefully to avoid the external contamination by hand. Write down the completion time of exposure.
- 4.2.7 Collect the SCDA plate in S.S. petricane and keep the petricane in dynamic Pass box of Microbiological testing area and follow the procedure for incubation.

4.3 Procedure for Incubation:



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4.3.1	Incubate SCDA plates in inverted position along with control					
1.3.1	25°C for 72 hours.	or place (unexposed) at 20				
4.3.2	Control plate represents the medium sterility.					
4.3.3	On completion of incubation transfer the same plates include	ling control plate for further				
4.3.3	incubation at 30°–35°C for 48 hours.	ang control plate for further				
4.3.4	After the completion of 05 days incubation period, observe the	e SCDA plates and count the				
4.5.4	bacterial and fungal colonies. Record the result in respective A	•				
4.3.5	Bacterial and fungal count should be within limit as per point N					
4.4	Precaution:	VO., 4.10				
4.4.1						
4.5	Active Air sampling:	it having no crack.				
4.5.1	Take filtered 70% v/v IPA and lint free sterile mopping cloth.					
4.5.2						
7.3.2	Prepare Soybean casein digest agar medium as per SOP and mark the plates with Name of Medium, Lot No., Date of Preparation, Use before and Sign and date on bottom of plates					
	and proceed for pre incubation not less than 48 hours at 20°–2:	•				
4.5.3	Take required number of preincubated plates	<i>J</i> .				
4.5.4	Keep the plates in petricanes, previously disinfect the petricans v	with 70 % v/v IPA and keen in				
4.5.4	the dynamic pass box.					
4.5.5	Follow the entry/exit procedure as per reference SOP for Micro	shiplogy togting area				
4.5.6	Disinfect the outer surface of air sampler with sterile mop, using					
	1					
4.5.7	Sterilise the S.S. perforated lid of air sampler by autoclaving testing area through dynamic pass box.	and transfer to inicrobiology				
150		data of compling with montron				
4.5.8	Before sampling, mark the sterile SCDA plate with location and	date of sampling with marker				
450	pen on the bottom of plate.	the starile CODA alete in the				
4.5.9	Open the lid of air sampler and put near the site of sampling. Plants and the site of sampling and put near the site of sampling.	•				
4.5.10	the sampling head of the instrument and close with perforated lid					
4.5.10	Operate the Air Sampler as per SOP" Operation and Cleaning of	•				
4.5.11	Sample 1m ³ (1000 litre) of air from each assigned location as per					
4.5.12 Ensure that the sample quantity is displayed on the screen of the instrument after each						



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	sampling operation.				
4.5.13	After Air sampling, open the perforated lid of Air sampler an	nd take out the SCDA plate, close			
	the lid and put in to the S. S. Petri canes/containers				
4.5.14	Disinfect the perforated lid by mopping with sterile mop u	sing 70% v/v IPA before taking			
	another sample.				
4.5.15	Collect the SCDA plate in S.S. petricane and keep the pe	tricane in dynamic Pass box of			
	microbiology testing area and follow the procedure for incul	bation.			
4.6	Procedure for Incubation				
4.6.1	Incubate SCDA plates in inverted position along with co	ntrol plate (unexposed) at 20°-			
	25°C for 72 hours.				
4.6.2	Control plate represents the medium sterility.				
4.6.3	On completion of above incubation transfer the same plates including control plate for				
	further incubation at 30°-35°C for 48hours.				
4.6.4	After the completion of incubation period, observe the SCDA plates and count the bacterial				
	and fungal colonies. Record the result in respective Annexus	re-II.			
4.6.5	Bacterial and fungal count should be within limit as per poir	nt No.: 4.10			
4.7	Surface monitoring:				
4.7.1	Perform the surface monitoring in the locations according	to the pre-defined locations and			
	surface monitoring schematic diagrams as per Annexure-IV				
4.7.2	Perform the surface monitoring by following methods.				
4.8	55 mm surface contact (RODAC) plates:				
4.8.1	Carefully open the RODAC plate, invert and contact the sur	face of agar to the surface to be			
	monitored. Press the plate firmly to expose the whole surfac	e of agar to be sampled surface.			
4.8.2	Slowly take back the plate and close the plate with lid. Take	care to not to leave any traces of			
	agar medium to the surface monitored				
4.8.3	Disinfect the sampled area and clean with sterile moping page	d			
4.8.4	After completion of test label each plate on bottom side with	h location code, shift, sign (initial)			
	and date.	. . ,			
4.8.5	Perform the sampling in all the locations as per the sampling	g plan using same procedure.			
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- 4.8.6 Place all the plates in SS carriers
- 4.8.7 Perform the monitoring at the end of activity (before cleaning).
- 4.8.8 Record the

4.10

- 4.9 Procedure for Incubation
- 4.9.1 Incubate SCDA plates in inverted position along with control plate (unexposed) at 20°–25°C for 72 hours.
- 4.9.2 Control plate represents the medium sterility.
- 4.9.3 On completion of above incubation transfer the same plates including control plate for further incubation at 30°–35°C for 48 hours.
- 4.9.4 After the completion of incubation period, observe the SCDA plates and count the bacterial and fungal colonies. Record the result in respective Annexure-III
- 4.9.5 Bacterial and fungal count should be within limit as per point No.: 4.10
- 4.9.6 **Trend Preparation:** Prepare the trend for the result on monthly basis of Passive air sampling test result and Quarterly for Active air sampling test result.

Active Air sampling (cfu/m3)						
		Bacter	ia	Fungus n Alert Limit Action Limit Specifi		
Grade	Alert Limit	Action Limit	Specification			Specification
Grade A #			<1			Nil
Grade B			10			Nil
Grade C			100			Nil
Grade D			200			Nil

Settle Plate (cfu/Plate)						
	Bacteria			Fungus		
Grade	Alert Limit	Action Limit	Specification	Alert Limit	Action Limit	Specification
Grade A #			<1			Nil
Grade B			5			Nil
Grade C			50			Nil
Grade D			100			Nil



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cfu/RODAC Plate							
			Bac	teria			Fungus
Grade	Alert Limit		Action Limit		Specif	fication	
	Wall	Floor	Wall	Floor	Wall	Floor	
Grade A #					< 1	< 1	Should be
Grade B					05	05	Absent.
Grade C					25	25	
Grade D					50	50	

#: Any count observed in any location in Grade A shall trigger investigation.

Note: Action and alert limit shall be specified after generating the graphical data for a period of one year for microbial test result by passive air sampling, active air sampling and surface monitoring.

4.12 Frequency:

- 4.12.1 Passive Air Sampling: Daily during operation
- 4.12.2 Active Air Sampling: Weekly during operation
- 4.12.3 Surface Monitoring: Once in Week at end of the operation.

5.0 ANNEXURE (S):

Annexure-I: Environmental Monitoring of Microbiology testing area report by Passive Air Sampling testing report.

Annexure-II: Environmental Monitoring of Microbiology testing area report by active air sampling.

Annexure-III: Environmental Monitoring of Microbiology testing area by surface monitoring.

Annexure-IV: Surface monitoring schematic diagrams for Environmental Monitoring of Microbiology testing area (Passive/Active/RODAC).

6.0 REFERENCE (S):

SOP: Entry and exit to the Microbiology Testing Area

SOP: Procedure for preparation of media.



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SOP: Investigation of out of specification of test result

SOP: Operation and cleaning of Air Sampler.

SOP: Preparation, Approval, Distribution control, revision and Destruction of Standard

operating Procedure (SOP).

7.0 ABBREVIATION (S) / DEFINITION (S):

LAF : Laminar Air Flow

S.S. : Stainless Steel

BOD : Bacteriological Oxygen Demand.

Cfu/m³: colony forming unit per cubic meter.

SCDA: Soyabean casein digest agar

v/v : volume/volume

IPA: Iso Propyl Alcohol

QC: Quality Control

QCM: Quality Control Microbiology

SOP: Standard Operating Procedure

REVISION CARD

S.No.	REVISION No.	REVISION DATE	DETAILS OF REVISION	REASON (S) FOR REVISION	REFERENCE CHANGE CONTROL No.
01	00			New SOP	



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Annexure 1

ENVIRONMENTAL MONITORING OF MICROBIOLOGICAL TESTING AREA BY PASSIVE AIR SAMPLING (SETTLE PLATE METHOD)

SOP Reference No.:

Date of sampling	:	Date of Report	:	
Sampling time	:	Sampled By (Sign. &Date)	:	
Area status	: In operation /At rest condition			

Media Details	Incubation	Incubation	Incubator ID No.
	Temperature	Time	
Soyabean Casein Digest Agar	20°–25°C	72 hours	
Lot No.:	30°-35°C	48 hours	

S.No.	Location ID. No.	Sampling Location Description	Grade	Count (C	Cfu / plate)
	1D. No.			Bacteria	Fungi
1.	S-01	Change room – I	D		
2.	S-02	Change room – I	D		
3.	S-03	Change room – II	С		
4.	S-04	Change Room – III	В		
5.	S-05	MLT Room	В		
6.	S-06	MLT Room	В		
7.	S-07	MLT Room	В		
8.	S-08	MLT Room	В		
9.	S-09	MLT Room	В		
10.	S-10	Change Room – IV	В		
11.	S-11	Upper side Laminar air flow	В		
12.	S-12	Right side under Laminar air flow	A		
13.	S-13	Left side under Laminar air flow	A		
14.	S-14	Dynamic pass box(media preparation room)	A		
15.	S-15	Dynamic pass box(Incubator room)	A		
16.	S-16	Garment Cubical	A		



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Frequency: Daily in operation	Negative control:
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Limit: Count (Cfu / plate)				
Grade	Alert limit	Action Limit	Specification limit	
Grade A			<01CFU	
Grade B			<05 CFU	
Grade C			<50 CFU	
Grade D			<100 CFU	

Remark: Area Complies / Does not Complies with specified limit.

Observed By	Date of Deposit	Checked By
(Sign. & Date)	Date of Report	(Sign. & Date)



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Annexure 2 ENVIRONMENTAL MONITORING OF MICROBIOLOGICAL TESTING AREA BY ACTIVE AIR SAMPLING

| Date of sampling | : | Date of Report | : | | Sampling time | : | Sampled By (Sign. & Date) | : | | Area status | : In operation / At rest condition |

Media Details	Incubation Temperature	Incubation Time	Incubator ID No.
Soyabean Casein Digest Agar	20°–25°C	72 hours	
Lot No.:	30°-35°C	48 hours	

S.No.	Location	Compling Location Description	Grade	Count (Cfu / plate)
5.110.	ID. No.	Sampling Location Description	Grade	Bacteria	Fungi
1.	A-01	Change room – I	D		
2.	A-02	Change room – II	С		
3.	A-03	Change Room – III	В		
4.	A-04	Change Room – IV	В		
5.	A-05	MLT Room Left side of LAF	В		
6.	A-06	MLT Room in front of change room IV	В		
7.	A-07	Right side under Laminar air flow	A		
8.	A-08	Left side under Laminar air flow	A		
9.	A-09	Dynamic pass box(media preparation room)	A		
10.	A-10	Dynamic pass box(Incubator room)	A		
11.	A-11	Garment Cubical	A		

Frequency: Daily in operation Negative control:

Limit: Count (Cfu / plate)				
Grade	Alert limit	Action Limit	Specification limit	
Grade A			<01CFU	
Grade B			<10 CFU	
Grade C			<100CFU	
Grade D			<200 CFU	

Remark: Area Complies / Does not Complies with specified limit.

Observed By:	Data of Danort	Checked By:
(Sign. & Date):	Date of Report	(Sign. & Date):



Date of sampling

Sampling time

PHARMA DEVILS

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Annexure 3 ENVIRONMENTAL MONITORING OF MICROBIOLOGICAL TESTING AREA BY SURFACE MONITORING (RODAC PLATE METHOD)

SOP Reference No.:

Date of Report :

Sampled By (Sign. & Date) :

Area status : In operation /At rest condition

:

:

Media Details	Incubation Temperature	Incubation Time	Incubator ID No.
Soyabean Casein Digest Agar	20°–25°C	72 hours	
Lot No.:	30°-35°C	48 hours	

S.No.	Location Compling Location Description		Grade	Count (Cfu / plate)	
5.110.	ID. No.	Sampling Location Description	Grade	Bacteria	Fungi
1.	R-01	Change room – I	D		
2.	R-02	Change room – II	С		
3.	R-03	Change Room – III	В		
4.	R-04	Change Room – IV	В		
5.	R-05	MLT Room in front of change –III	В		
6.	R-06	MLT Room in front of change –IV	В		
7.	R-07	MLT Room (middle of the room)	В		

Frequency: Weekly Negative control:

Limit: Count (Cfu/plate)					
Grade	Alert limit	Action Limit	Specification limit		
Grade B			<05 CFU		
Grade C			<25 CFU		
Grade D			<50 CFU		

Remark: Area Complies / Does not Complies with specified limit.

Observed By	Date of Report	Checked By	
(Sign. & Date)	Dute of Report	(Sign. & Date)	



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Annexure 4 MICROBIOLOGY ENVIRONMENT MONITORING LOCATION DIAGRAM