

PHARMA DEVILS MICROBIOLOGY DEPARTMENT

		STANDARD OPERATING PROCEDURE	
Departn	nent: Microbiology		SOP No.:
Title: M	edia Fill		Effective Date:
Superse	des: Nil		Review Date:
Issue Da	ite:		Page No.:
1.0	OBJECTIVE	: To lay down a procedure for Media Fill o Filling facility.	peration in the sterile dry powder
2.0	RESPONSIBILITY	: Operator concerned/Production officer/Q.	A Officer
3.0	ACCOUNTABILITY	: Production Manager/QC Manager.	
4.0	PROCEDURE	:	
4.1	PRE-START UP:		
4.2	Ensure that all the equipme validated.	ents, HVAC system, water system and other uti	lity services of the facility are
4.3		adiation certification report of the lactose + SC and sterility test report (done in-house) is avail	
т.,	•	eport and growth promotion/inhibition test repo bility should be NLT 1gm/10 ml of WFI].	ort of sterile lactose + SCDM
4.4	Ensure that freshly distilled	WFI to be used for filling, is autoclaved and t	he sample given for sterility test.
4.5	Ensure that all the contact j	parts of Dry Powder Filling machine are duly c	leaned and sterilized or sanitized.
	Ensure that the last Environ	nmental Control Reports of the area are conform	ming to the acceptance standard.
4.6	Ensure that the Liquid Filli autoclaving /sanitization.	ng machine is done inside the sterile filling are	a on the previous day after



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4.9	Ensure that the cleaning, sanitization and fumigation of th	e area is done on the previous day as per SOP.		
	Ensure by manometer readings, that the pressure balancing	g of the Sterile area is as per requirement		
5.0	START UP			
5.1	Enter the sterile area as per SOP using gowns sterilized 4	days back.		
5.2	Check the cleanliness of the Sterile area.			
5.3	Check the cleanliness of the Powder Filling machine.			
5.4	Check and confirm the temperature and RH of powder filling room is as per the requirement.			
5.5	Assemble the Powder Filling machine. Ensure that that fi simultaneously.	lling of powder and liquid can be done		
5.6	Measure the nonviable particulate count of filling cabinet	and filling room		
6.0	OPERATION:			
6.1	Bring the autoclaved WFI container near the liquid filling	assembly.		
6.2	Transfer sterile lactose + SCDM powder mixture from the	e container in to powder hopper.		
6.3	Transfer sterile dried rubber stoppers (sterilized 4 days bases and sample for sterility testing simultaneously.	ck) in to the hopper of stoppering unit. Send		
6.4	Put the inlet suction tube of Liquid Filling Assembly into	the WFI container.		
6.5	Set the Powder Filling machine for the respective dose.			



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8.3	Remove the Liquid Filling Assembly and all the contact parts. Disterilized/sanitised.	isinfect it, clean it and get it
8.4	Ensure that all the left over rubber stoppers are given outside the	sterile area for destruction.
8.5	Shut down the vial powder filling machine as per SOP.	
8.6	CLEANING:	
8.6.1	Ensure that all the tools and accessories containers etc. used durin cleaning/sanitization/sterilisation.	g media fill are given for
8.6.2	Follow the cleaning and sanitization SOP for cleaning of sterile and	rea.
8.6.3	Do the extra cleaning and sanitization of the floor/walls/machine a Bacillocide and Lysol solution.	after media fill run with 10%
9.0	ACCEPTANCE CRITERIA:	
9.1	Initial validation: During initial validation, it should qualify all t during each run there should not be growth in more than two vials	
9.2	Revalidation: Only one media fill run in which there should not	be any growth in more than two vials.
10.0	ELIGIBILITY CRITERIA FOR PERSONNEL PERFORM	IING MEDIA FILL RUN:
10.1	Persons involved in media fill should be medically examined and	declared fit within last one year.
10.2	Persons should be trained on general hygiene and current gowning condition should be O.K.	g procedure, and the present health

10.3 The personnel should be microbiologically monitored during run.



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11.0	NOTES	
11.1.0	In case of any positive growth in any vial during the incubation per	riod.
11.1.1	It should be isolated and identified to the genus level.	
11.1.2	If the isolated organism is other than the house flora, thorough investig Quality Assurance and Production.	ation shall be carried out by
11.1.3	The source of the contamination must be established.	
11.2	In case of any failure in media fill run:	
11.2.1	When there is no assignable cause, media fill shall be repeated three tir commence only after all the three runs meet the acceptance criteria.	nes and production should
11.2.2	When there is assignable cause, after rectification of the cause, repeat t	he media fill run once.
11.3	Before and during media fill run no special cleaning shall be carried ou	ıt.
11.4	Normal production can resume only after minimum one day of environ report.	mental monitoring compliance
11.5	Batches filled before the final result of the media fill run shall not be remedia fill run passes in case of initial validation.	cleased to the market till the
11.6	The routine environmental monitoring plates shall be kept for 14 days investigation of any positive growth in the media filled vials. If for any considered invalid vials shall not be incubated.	
11.7	Media fill runs can be aborted for the same reason that a product lot we units filled before an incident that would cause an aborted fill must be	
11.8	During media fill all the Operators, Officers & Maintenance staff who filling, supervision and maintenance must be involved in media fill tria	



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- **11.9** The volume of liquid filled must be sufficient to wet all surfaces including the closure and to facilitate inspection.
- **11.10** The line must be run at a slower speed than normal production run to give greater exposure time.
- **11.11** Total duration of the routine media fill must be the same or more as the longest process conducted on that line.
- **11.12** The incubation temperature of the filled containers must be sufficient to promote microbial growth at 30-35°C. for 168 hrs. (stored upright) followed by 20-25°C. for 168 hrs. (stored upside down).
- **11.13** Personnel that conduct the inspection of incubated media fills must have training on basic microbiological concepts, concepts of media fill and examples of contaminated container showing various stages of growth.

12.0 POST MEDIA FILL RUN:

- **12.1** Filled incubated vials should be optically checked by microbiologist and certified.
- **12.2** Incubate the vial samples with no growth approximate 5 vials for 14 days with normal house flora and 5 vials with organisms used for the sterility test growth promotion for 14 days.
- **12.3** Temp. range for incubation 1st week 30 to 35°C. (vials stored upright). 2nd week 20 to 25°C.(vials stored upside down). These vials should show promotion of growth.



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ANNEXURE - I COMBINATION OF VIAL AND RUBBER STOPPER.

S.No.	VIAL		RUBBER STOPPER			
	VIAL SIZE	ТҮРЕ	GREY BUTYL	HALO BUTYL		
1.	5 ML	III	_/			
2.	7.5 ML	III	_/			
3.	10 ML	III	_/			
4.	15 ML	III	_/			
5.	20 ML	III	_/			



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ANNEXURE - II ACCEPTANCE CRITERIA FOR MEDIA FILL SIMULATION

MAXIMUM NO.OF REJECTED VIALS ACCEPTABLE AT	PERCENTAGE OF FAILURE	BATCH SIZES					
95% CONFIDENCE LEVEL		5000	10000	20000	50000	100000	INFINITY
0	0.1%	2469	2668	2808	2912	2951	2995
1	0.1%	3676	4047	4339	4575	4670	4747
2*	0.1%	4684	5207	5670	6044	6207	6294
0	0.01%				24698	26686	29944
1	0.01%				46093	47047	47047
2	0.01%					62911	62911