

PHARMA DEVILS QUALITY ASSURANCE DEPARTMENT

Document Name: Performance Qualification test datasheet # 3.3 for Lyophilizer					
Equipment/System ID:		Document Number:			
Effective Date:		Version Number: 00			
Test ID #3.3: Cleaning In Place Performance check and SIP Validation		SIP Validation	Test Run: _		
Target: Necessary	 The objective of this test is to verify the Cleaning in performance check within the chamber is adequate, as per system specification. The objective of this test is to verify that all locations within the chamber and in the condenser are exposed to the sterilization temperature during SIP cycle. The chamber and condenser attaining a temperature of 122°C during the sterilization hold (For 30 min.) period with steam pressure of 2.11 to 2.33 bar A. (This is considering the absolute pressure default value of 1.01 Bar) Temperature spread within the range of 122°C to 124.5°C during sterilization cycle will demonstrate the uniform heat distribution within the chamber. Any location(s) where the temperature sensor is placed, achieving minimum sterilization temperature throughout the sterilization hold period will be considered as cold spot. Calibrated Data logger 				
materials:	 Calibrated T-type temperature sensor Lyophilizer in Auto mode 				
Preconditions:	 Equipment should be normal operation The chamber should be empty. The temperature of the chamber should Ensure that Steam quality tests perform 	al mode d not be more than 20°C from ambient ned and result complies as per specific	ations.		
Test ID	More than Test Description				
1	Before SIP validation study ensures that t	he lyophilizer is at ambient Temperatu	ure.		
2	Take the calibrated data logger with minimum number of calibrated probes considering 5 probes for top and bottom shelf and 3 probes per shelf and one probe for the drain point.				
3	Prepare at least 49 Nos. calibrated temperature mapping probe with location and channel tag.				
4	Perform full loop calibration (probe + data logger) before starting the qualification runs. Attach calibration reports. If external agency is carrying out the test the calibration certificate will be referred.				
5	Pass 49 nosof Temperature mapping p Lyophilizer. Seal the port with silicon Suspend the probes in the chamber in dif surface. Place all sensors, biological in Drawing No 1 and Drawing No. 2 in using a strip of autoclavable tape, these data is recorded. Suspended the probes a chamber in different position so that p position of the probes and the biological in	probe into chamber through the valid the sealant so that steam leakage does ferent position so that probes do not t dicator and chemical indicator as p Appendix 3.3.1 . Label the thermocou- long with the self contained biological robes do not touch any metallic sur- ndicators in a representative schematic	lation port of the s not take place. ouch any metallic er the mentioned ouples by number ple for which the l indicators in the rface. Record the c form.		
6	Keep one probe at drain point of lyophilizer chamber. The drain point is monitored because to identify the coolest point during the cycle.				
7	Set the standard recipe for SIP cycle.				
8	Record the parameter of recipe in the data	asheet.			
9	Start the data logger to record actual temp to time at a scan rate of 5 sec.	peratures within the sterilization cham	ber with respect		



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10	Run the cycle as per the SOP and observe	e the temperature on SCADA and data	logger		
11	When the SIP cycle completes; (1) Collect thermograph from the multipoint temperature recorder of the Lyophilizer and (2) Download the data from data logger into the computer for data-analysis and printing. Record the temperatures observed at different locations (3) remove and collect the exposed biological indicators and send them within a self-sealing sterile poly bag to microbiology laboratory for further studies with necessary identification.				
12	Compare the lyophilizer report with the recipe feed.				
13	Calculate Fo value of each temperature mapping location.				
14	After the cycle is over visually check the chamber and condenser for the dryness.				
	Identify the coolest point and calculate the mathematical F0 value for the coolest point observe during the cycle as per the formula below. For calculation of F0 value the temperature for th cold point is to be recorded in the datasheet for each one-minute time interval at the hold time of sterilization.				
15		$F0 = dt \sum 10^{\left\lfloor \frac{T-121}{Z} \right\rfloor}$			
	F0 = dt Σ (lethality rate) Where, dt = the time interval between successive temperature measurements (1 min.). T = the observed temperature at that particular time (As per the actual temperatures recorded) Z = the change in the heat resistance of Geobacillus Stearothermophilus spores as temperature is changed (10°C)				
16	If the study is acceptable perform two more consecutive replicate runs to demonstrate cycle and sterilization-in-place reproducibility.				
17	Compile the data generated during the qualification test for evaluation of the system.				
18	Record the observations & data as per Appendix 3.3.1.				
19	Attach the print out of the test cycle taken from the Lyophilizer with this data sheet.				
	Acceptance	criteria	Acceptance criteria fulfilled? (Y/N)		
1.	For Chamber and Condenser				
1.1	Each external probe should achieve a min sterile hold for NLT 30 minutes	nimum temperature of 122°C at			
1.2	All inbuilt probes should achieve a minin hold and run for the set time.	num temperature of 122°C at sterile			
1.3	No external probe should vary by more the probe temperature during the sterilization	han 1° C from average of all external hold time.			
1.4	The temperature spread at the sterilization vary by more than 2°C.	n hold time for each probe should not			
1.5	The external drain probe temperature sho the inbuilt drain probe temperature at the	uld not vary by more than 2° C from sterilization hold time.			



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1.6	The lag time should not more than 30 sec	S.				
1.7	The mathematical F0 value calculated for the coolest point should be more than the Biological F0 value for the indicators or 30 minutes.					
1.8	Chemical indicators at all locations should change the color from pink to green for at least three compartments.					
1.9	All BIs should show no growth after incubation of 55-60°C for 48 hours and 12 log reductions should be achieved.					
1.10	The report generated should be in similar sequence as per the recipe feed.					
1.11	The cycle should run in a similar sequence as per the recipe.					
1.12	The condenser and chamber should be visually dry enough.					
2.	For In Line Air Filter					
2.1	Visually inspect the Filters for drying.					
2.2	The minimum temperature at each location should be ≥ 122.0 °C during sterilization hold for NLT 30 minutes					
2.3	The F0 Value for all the probes should not less than the biological Fo Value or 30 minutes, whichever is greater					
2.4	Chemical indicators at all locations should change the color from pink to green for at least three compartments					
2.5	All BIs should show no growth after incubation of 55-60°C for 48 hours and 12 log reductions should be achieved					
3.	For Thermocouples placed outside chamber and condenser in the ninelines.					
3.1	The minimum temperature at each location should be ≥ 122 °C during sterilization. Hold period NLT 30 minutes					
3.2	The F0 Value for all the probes should not less than the biological Fo Value or 30 Minutes, whichever is greater					
3.3	Chemical indicators at all locations should change the color from pink to					
3.4	All BIs should show no growth after incubation of 55-60°C for 48 hours and 12 log Reduction to be achieved.					
Measures after test execution:	Allow all the sensors to cool to 65°C, before starting the next cycle.					
Comment Ref. No.	Comment		Deviation Ref. No.			
Checked by (Signature/ Date)		Verified by (Signature/Date)				